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A Brain-Machine-Brain Interface for Rewiring of Cortical Circuitry after Traumatic Brain Injury

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Introduction

The goal of this project is to use an implantable brain-machine-brain interface to enhance behavioral recovery after traumatic brain injury (TBI) by reshaping long-range intracortical connectivity patterns. We hypothesize that artificial synchronous activation of distant cortical locations will encourage spontaneously sprouting axons to migrate toward and terminate in the coupled region, and that such directed sprouting can aid in functional recovery.

Body

In this section of the annual report, we describe the research accomplishments associated with each task outlined in the approved Statement of Work.

1. Electronics Development

For **Tasks 1.1** and **1.2**, A fully integrated neural recording front-end comprising a low-noise two-stage amplification circuitry and a 10-bit successive approximation register (SAR)-based analog-to-digital converter (ADC) was designed and fabricated using the AMS 0.35 μ m 2P/4M CMOS process. The ac-coupled amplification circuitry provided a maximum mid-band ac gain of ~66 dB at 1 kHz and featured a measured input-referred noise voltage of ~3.1 μ V_{rms} from 0.5 Hz to 50 kHz, while dissipating ~27 μ W from 1.5 V. The SAR ADC featured an effective number of bits (ENOB) of 9.2 for sampling frequency of 35.7 kS/s, while dissipating only 6 μ W. Monolithic circuitry was designed using standard digital cells to identify the presence of large action potentials in the recorded digitized data with a spike discrimination algorithm based on two programmable threshold levels and time-amplitude windows. A first-order digital highpass filter was also designed to remove any dc/low-frequency artifacts prior to spike discrimination. The measured power consumption per channel was less than 1 μ W with a 1.5-V supply and ADC sampling frequency of 35.7 kS/s.

For **Task 1.3**, a template subtraction algorithm for real-time stimulus artifact rejection (SAR) based upon an infinite impulse response (IIR) temporal filtering technique was developed. We demonstrated that initialization of the memory within the system architecture using the first recorded stimulus artifact could significantly decrease the system response time as compared to the case without memory initialization. Using two sets of prerecorded neural data from an *Aplysia californica*, the algorithm was shown in simulation to be capable of eliminating virtually all the stimulus artifacts in real time and recovering the extracellular neural activity with μ W-level power consumption from 1.5 V.

For **Tasks 1.4** and **1.5**, a decision-making circuitry was implemented as a programmable gate array to provide any logic combination of the four spike discriminator outputs (SDO 1~4) as a trigger signal for stimulation activation using a 16b combination code for each stimulation channel. A high-output-impedance current microstimulator was also designed and fabricated using the AMS 0.35 µm 2P/4M CMOS process. It delivered a maximum current of 94.5 µA to the target cortical tissue with current efficiency of 95.6% and voltage compliance of 4.68 V with a 5-V supply, when configured for monophasic stimulation with passive discharge. The programmable microstimulator could also be configured to deliver charge-balanced asymmetric biphasic stimulus with programmable time/amplitude parameters. The stimulus current could be programmed via a 6-bit digital-to-analog converter (DAC) with accuracy better than 1.6 LSB.

For **Task 1.6**, a highly integrated voltage converter was designed and fabricated using the AMS 0.35 μ m 2P/4M CMOS process to generate a 5-V supply for the stimulating back-end from a miniature primary battery that powers the entire system. The voltage converter employed only one external capacitor (1 μ F) for storage, and delivered a maximum dc load current of ~88 μ A with power efficiency of 31% with its output voltage adjusted to 5.05 V. This current drive capability could afford simultaneous stimulation on all eight channels of the system with current amplitude up to ~100 μ A and average stimulus rate >500 Hz, which is comfortably higher than firing rate of cortical neurons (up to 150 spikes per second).

For **Task 1.7**, all electronic circuitry (except for that related to **Task 1.3**) was fabricated using the AMS 0.35 μ m 2P/4M CMOS process, measuring 3.3 × 3.3 mm². Details of circuit and system architectures, electrical performance characterization, and *in-vivo measurement* results were reported in Appendix I (Azin et al., *IEEE J. Solid-State Circuits*, pp. 731-745 April 2011).

2. Neurobiology

As documented in the annual report submitted by the Partnering PI, Prof. Randolph Nudo, for Award W81XWH-10-1-0742, substantial progress has been made in the Neurobiology tasks. In addition to obtaining all the required approvals from the local Institutional Animal Care and Use Committee (IACUC) and ACURO for the rodent studies, we are now able to drive temporal coupling of activity from the rostral forelimb area (RFA) to a distant cortical location, such as the caudal forelimb area (CFA), primary somatosensory forelimb area (S1 forelimb area), and second somatosensory field (SII), in a rat model in both acute and chronic settings.

Activity-dependent stimulation (ADS) from the RFA to the S1 forelimb area has also been performed in brain-injured rats. As can be seen in Partnering PI's report, the results demonstrated an unprecedented, potent effect of ADS on motor performance after only 5 days of operation. By Day 14 post-lesion, performance in the ADS group was indistinguishable from pre-lesion performance (\sim 70% successful retrievals). In the ADS group, the mean percent pellets retrieved increased from \sim 20% 3 days post-lesion to \sim 40% 5 days post-lesion and >50% 8 days post-lesion. Statistical analysis is currently being completed by including more animals in the study. Detailed description of the methods used and quantitative analysis of the results obtained can be found in Partnering PI's annual report.

Key Research Accomplishments

- Design and fabrication of integrated neural recording front-end incorporating amplification and digitization.
- Demonstration of *in-vivo* functionality of front-end by recording neural spikes from cerebral cortex of anesthetized rat.
- Develop and test a template subtraction-based neural signal-processing algorithm for real-time stimulus artifact rejection.
- Design and fabrication of integrated neural microstimulator.
- Demonstration of *in-vivo* functionality of microstimulator by driving neck movement after stimulating primary motor cortex of anesthetized rat.
- Design and fabrication of very-low-power digital filtering and spike discrimination circuitry.
- Design and fabrication of highly integrated dc-dc converter to generate stimulation power supply of 5 V from a single 1.5-V battery.

Reportable Outcomes

1- Manuscripts/Abstracts/Presentations:

- P. Mohseni, "Wireless integrated systems for chemical and electrical neural recording: Challenges in high-site-density brain monitoring," *RF Biomedical Electronics and Sensors Workshop*, *Int. Microwave Symp. (IMS'11)*, Baltimore, MD, June 5, 2011.
- P. Mohseni, "Toward closing the loop: Circuits and systems for real-time neurochemical sensing and activity-dependent intracortical microstimulation," *Berkeley Sensor and Actuator Center (BSAC) Seminar Series, University*

- of California-Berkeley, Berkeley, CA, April 26, 2011.
- M. Azin, D. J. Guggenmos, S. Barbay, R. J. Nudo, and P. Mohseni, "A battery-powered activity-dependent intracortical microstimulation IC for brain-machine-brain interface," *IEEE J. Solid-State Circuits*, vol. 46, no. 4, pp. 731-745, April 2011.
- 2- Patents and Licenses Applied for/Issued: None yet.
- 3- Degrees Obtained from Award: Ph.D. (Meysam Azin), May 2011
- 4- Development of Cell Lines and Tissue/Serum Repositories: Not applicable.
- 5- Infomatics (Databases and Animal Models): None yet.
- 6- Funding Applied for: None yet.
- 7- Employment/Research Opportunities Applied for/Received: Senior Engineer (Meysam Azin), QualComm

Conclusion

In conclusion, rapid progress is being made toward developing smart prosthetic platforms for altering plasticity in the injured brain, leading to future therapeutic interventions for TBI that are guided by the underlying mechanisms for long-range functional and structural plasticity in the cerebral cortex. Our first-generation integrated device was tested successfully in a rat model in both acute and chronic settings by recording neural spikes from one cortical region and subsequently driving microstimulation of a distant cortical region with a user-adjustable spike-stimulus time delay. Using a controlled cortical-impact device, we have successfully induced TBI in the CFA, sparing the RFA. Behavioral assessments of reaching, retrieval of small food items, and locomotion demonstrate that deficits persist during the 5-week recovery period following injury. Further, an unprecedented, potent effect of ADS on motor performance after only 5 days of operation has been demonstrated in rats with TBI. Statistical analysis is currently being performed by including more rats in the study and creating a second control group of brain-injured rats that receive intracortical microstimulation independent of the neural activity in the RFA. We envision having the results ready in Q1 of Year 2.

Appendix I M. Azin, D. J. Guggenmos, S. Barbay, R. J. Nudo, and P. Mohseni, "A battery-powered activity-dependent intracortical microstimulation IC for brain-machine-brain interface," *IEEE J. Solid-State Circuits*, vol. 46, no. 4, pp. 731-745, April 2011.

A Battery-Powered Activity-Dependent Intracortical Microstimulation IC for Brain-Machine-Brain Interface

Meysam Azin, Student Member, IEEE, David J. Guggenmos, Scott Barbay, Randolph J. Nudo, and Pedram Mohseni, Member, IEEE

Abstract—This paper describes an activity-dependent intracortical microstimulation (ICMS) system-on-chip (SoC) that converts extracellular neural spikes recorded from one brain region to electrical stimuli delivered to another brain region in real time in vivo. The 10.9-mm² SoC incorporates two identical 4-channel modules, each comprising an analog recording front-end with total input noise voltage of 3.12 $\mu V_{
m rms}$ and noise efficiency factor (NEF) of 2.68, 5.9- μ W 10-bit successive approximation register analog-to-digital converters (SAR ADCs), 12.4- μ W digital spike discrimination processor, and a programmable constant-current microstimulating back-end that delivers up to 94.5 μ A with 6-bit resolution to stimulate the cortical tissue when triggered by neural activity. For autonomous operation, the SoC also integrates biasing and clock generation circuitry, frequency-shift-keyed (FSK) transmitter at 433 MHz, and dc-dc converter that generates a power supply of 5.05 V for the microstimulating back-end from a single 1.5-V battery. Measured results from electrical performance characterization and biological experiments with anesthetized rats are presented from a prototype chip fabricated in AMS 0.35 μ m two-poly four-metal (2P/4M) CMOS. A noise analysis for the selected low-noise amplifier (LNA) topology is presented that obtains a minimum NEF of 2.33 for a practical design given the technology parameters and power supply voltage. Future considerations in the SoC design with respect to silicon area and power consumption when increasing the number of channels are also discussed.

Index Terms—Activity-dependent microstimulation, brain-machine-brain interface, intracortical microstimulation, neural recording, neurostimulation, spike discrimination, system-on-chip.

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Color versions of one or more of the figures in this paper are available online at http://ieeexplore.ieee.org.

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I. INTRODUCTION

N THE PAST decade, significant advancements have occurred in developing engineered devices that bidirectionally interact with the central and peripheral nervous system. Specifically, brain-machine interfaces (BMIs) have been developed for human application that use intrinsic neuronal signals from the brain as input commands to control external devices. Cortical control of the movement of a prosthetic arm or mouse cursor on a computer screen using brain electrical signals have indeed been successfully demonstrated in non-human primates and even in case studies in humans suffering from high-level spinal cord injury [1]-[3]. Other types of BMIs use device-generated electrical signals for stimulation of the nervous system to modulate a particular neural pathway that might be dysfunctional, or to transmit a specific type of sensory information. Deep brain stimulation (DBS), a clinically effective neuromodulation therapy for Parkinson's disease (PD), and neuroprostheses to restore lost sensory functions in blind and hearing-impaired patients fall into this category [4], [5]. Neuroprostheses to restore a lost cognitive function such as memory are also on the horizon [6].

The emergence of BMI technology has been in part enabled by technological innovations in integrated sensor and circuit design over the past few years. Advancements in micro-electro-mechanical-system (MEMS) technology have produced millimeter-sized arrays of microelectrodes containing as many as 100 recording sites that allow long-term, reliable, and stable recording of neural signals [7], [8]. Similarly, novel design methods in application-specific integrated circuit (ASIC) development have recently led to numerous examples of low-power, low-noise, high-performance ICs for recording brain electrical activity [9]–[14], on-chip processing of the recorded neural data [15], [16], and stimulating the nervous system [17]–[21].

While existing BMIs continue to show great promise to improve the quality of life in people suffering from neuropathologies, a new generation of neuroprostheses is now emerging that aims to combine neural recording, signal processing, and microstimulation functionalities in a single device for closed-loop operation. This new category of neuroprostheses can use information extracted from brain neural activity to trigger microstimulation or modulate stimulus parameters in real time, potentially enhancing the clinical efficacy of neuromodulation in alleviating pathologic symptoms or restoring lost sensory and motor functions in the disabled. For example, it has been suggested

that the efficacy of DBS for PD can be increased by optimizing stimulation waveform parameters using a closed-loop global optimization algorithm [22], or that stimulation of the spinal cord directed by cortical neural activity can evoke or facilitate intended movement in the upper limb of paralyzed patients with spinal cord injury [23].

To that end, two discrete board-level devices were recently developed using commercially available hardware to provide refined sensory inputs in closed-loop neuroprosthetic systems [24] and induce neuronal plasticity for functional reorganization in an intact nervous system [25]. Both devices were proven functional in biological tests with awake rodents and primates, respectively. Further, Denison and colleagues have developed an implantable closed-loop microstimulator targeted at titration therapy in DBS [26]. The system incorporates a custom IC for multichannel sensing and analog spectral analysis of local field potentials (LFPs), an auxiliary microprocessor for key biomarker extraction and titration control, and a neurostimulation unit from an existing approved device. Liu and colleagues have proposed another closed-loop microstimulator, incorporating a biomedical multiprocessor system-on-chip (SoC) for real-time spike sorting with principal component analysis and a digitally controlled current stimulating back-end [27]. However, the sensing and digitization front-end is not co-integrated with the rest of the system. Further, neither of these works has yet been tested in vivo.

Finally, Flynn and colleagues have reported a 64-channel closed-loop DBS IC, incorporating eight channels of amplification for neural recording and an 8-bit logarithmic analog-to-digital converter (ADC) for digitization [28]. Similarly, Genov and colleagues have recently reported a single-chip 128-channel system that can be configured for fully differential recording and digitization of neural data as well as voltage-mode monophasic or biphasic stimulation on some or all electrodes [29]. However, in the absence of any on-chip neural signal processing unit, the stimulator operation in these works is not dependent on the recorded neural activity, and therefore system operation is not truly closed-loop.

We have previously reported an activity-dependent intracortical microstimulation (ICMS) chip that was limited to single-channel operation [30]. In this paper, we describe a single-chip battery-powered IC with autonomous operation for multichannel brain-machine-brain interface (BMBI) that converts extracellular neural signals recorded on one microelectrode to electrical stimuli delivered via another electrode in real time in vivo. The paper is organized as follows. Section II presents the proposed system architecture, whereas Section III discusses the design of major circuitry within the system. Section IV presents detailed benchtop measurement results characterizing electrical performance as well as in vivo results from biological experiments. Section V provides a noise analysis for the recording front-end and discusses future considerations in system design. Finally, Section VI draws some conclusions from this work.

II. SYSTEM ARCHITECTURE

Fig. 1 shows the proposed architecture for a proof-of-concept system along with its timing operation. There are two identical 4-channel modules per chip, each comprising a recording front-

end, monolithic digital signal processing (DSP) unit, and stimulating back-end. The recording front-end provides ac amplification, dc input stabilization, bandpass filtering, and 10-bit digitization to the recorded neural signals with fully programmable gain and bandwidth. The DSP unit provides additional high-pass filtering with an infinite impulse response (IIR) digital filter to remove any residual dc offsets or low-frequency artifacts, and performs real-time spike discrimination based on threshold crossing and two user-adjustable time-amplitude windows for all four channels in a fraction of one ADC conversion cycle. The accepted action potentials trigger the programmable stimulating back-end with a user-adjustable time delay (<29 ms) to deliver a charge-balanced asymmetric biphasic stimulus [19] or monophasic stimulus with passive discharge [18] to the neural tissue. Anodic and cathodic current pulse amplitudes can be programmed from 0 to 94.5 μ A and 31.5 μ A, respectively, with 6-bit resolution.

A decision-making circuitry implemented as a programmable gate array provides any logic combination of the four spike discriminator outputs (SDO 1–4) as a trigger signal for stimulation activation using a 16-bit combination code for each stimulation channel. Multiple recording and stimulation channels in conjunction with the adjustable time delay between spike discrimination and stimulus onset allows various input-output combinations and temporal relationships to be tested to enable *patient-specific* system operation for optimized performance.

The system can operate autonomously from a single 1.5-V battery. An on-chip dc-dc converter generates 5 V from the battery for increased voltage compliance in the stimulating backend. The dc-dc converter has similar architecture to those in [31], [32] and can provide an output voltage up to approximately $4 \times V_{DD}$ to a 1- μ F external capacitor for storage. A 735-bit latched shift register provides all programming and control bits for system operation, and a supply-independent relaxation oscillator with similar architecture to that in [33] generates the system clock with 5-bit programmable frequency for timing management.

A radio-frequency frequency-shift-keyed (RF-FSK) transmitter at 433 MHz is shared between the modules and can operate in two modes. In mode I, it transmits the full voltage record (raw or high-pass filtered) on one channel together with the corresponding spike discrimination event. A 3-bit preamble is used in each data stream for synchronization between the on-chip transmitter and an external receiver. Information bits are scrambled within the data stream to reduce the probability of preamble misdetection. In mode II, it transmits the spike discrimination events on all eight channels along with a 6-bit preamble in each data stream.

The system is designed to operate with a nominal clock frequency of 1 MHz. As illustrated in Fig. 1, each ADC conversion cycle takes 28 clock cycles in which 16 clock cycles are dedicated to sampling, 1 cycle to hold, 10 cycles to approximation and 1 cycle to reset, thus providing a sampling frequency of 35.7 kS/s/ch that is adequate for recording extracellular neural spikes with frequency components up to 10 kHz. The extended sampling period helps relax current drive requirement of the preceding ADC driver for low-power operation. The processing of

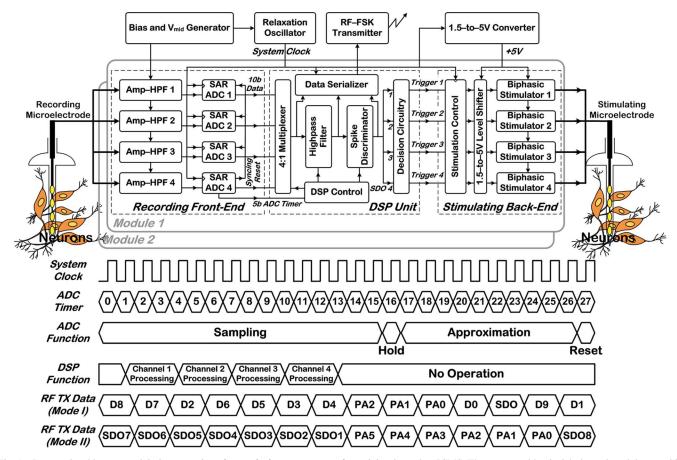


Fig. 1. Proposed architecture and timing operation of a proof-of-concept system for activity-dependent ICMS. There are two identical 4-channel modules per chip powered by a single 1.5-V battery.

each data channel in the DSP unit takes only three clock cycles, enabling the same DSP architecture to handle up to nine neural recording channels per module in any future expansion of system functionality.

III. INTEGRATED CIRCUIT ARCHITECTURE

A. Recording Front-End

Fig. 2 shows the circuit schematic of the analog recording front-end per channel, operating from 1.5 V and comprising a low-noise amplifier (LNA) with programmable bandwidth, G_m-C high-pass filter (HPF) with adjustable low cutoff frequency, and secondary amplifier with programmable gain and offset. The LNA provides a gain of 32 dB using capacitive feedback in conjunction with a pMOS pseudo-resistor to stabilize the dc input and provide a dc feedback path [14]. Two additional pMOS transistors have been used to short the LNA inputs and output during stimulation and prevent its saturation by large stimulus artifacts, if present. The LNA bandwidth can be programmed from 5.1 to 12 kHz by varying the tail current of the differential input stage. An ultralow-transconductance operational transconductance amplifier (OTA) and a 16.7-pF capacitor form a first-order HPF with an adjustable low cutoff frequency from 0 to 525 Hz by varying the OTA tail current. The analog HPF is intended to attenuate 60 Hz and remove low-frequency LFPs that might be present in the recorded neural data. The secondary amplifier with a resistive feedback provides additional gain of 19.9, 25.4, 27.9 and 33.6 dB, and has a 3-bit offset

adjustment mechanism by injecting a small dc current into the feedback path. The secondary amplifier provides a rail-to-rail output voltage swing and drives the \sim 12-pF capacitor network of the subsequent ADC that converts the amplified neural data to a 10-bit digital code for further processing by the DSP unit.

For analog-to-digital conversion in the recording front-end, a successive approximation register (SAR) ADC is used due to its low power consumption with similar architecture to that in [34]. The ADC design is modified as compared to that previously reported in [35] in order to operate with 1.5 V for lower power consumption.

B. DSP Unit

Fig. 3 shows the architecture of the DSP unit operating from 1.5 V, which is shared among all four channels in each module. Prior to spike discrimination, a first-order IIR filter provides additional high-pass filtering to the multiplexed digitized neural data with a programmable cutoff frequency of 366 or 756 Hz from 1-MHz system clock. Filter coefficients are selected to perform the filtering using arithmetic shifts, subtraction and addition only, with no need for digital multipliers or dividers. The operation of the time-amplitude window discriminator is also shown in Fig. 3 using neural spikes recorded wirelessly by our system from the cerebral cortex of a rat's brain. Once a filtered digitized sample crosses a user-positioned threshold level (L_0) , the spike discriminator will check whether the signal waveform

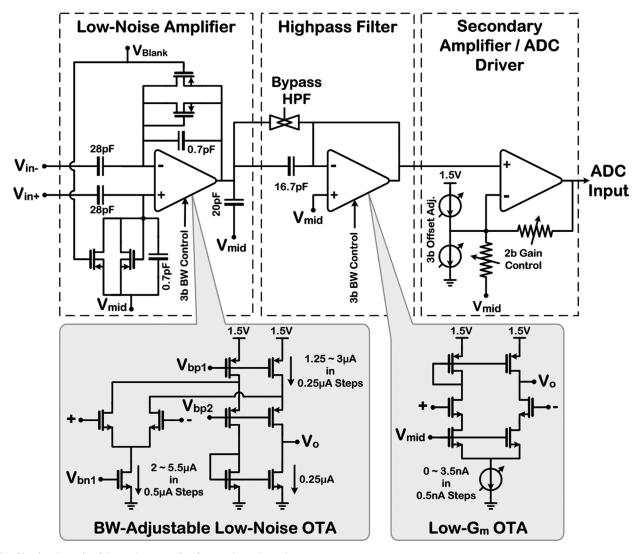


Fig. 2. Circuit schematic of the analog recording front-end per channel.

subsequently passes through two user-adjustable time-amplitude windows (solid red boxes). If a spike event is accepted on any channel, the corresponding SDO is activated after a programmable delay of 0 to \sim 28.6 ms for the decision circuitry to generate the stimulus trigger signal. The DSP operation is also blanked to disregard any incoming stimulus artifacts. Spike waveforms crossing the positive threshold level (L_0) are timealigned and superimposed in Fig. 3. The accepted spikes (dark grey) also pass through the two time-amplitude windows. Waveforms in light grey are rejected by the system (i.e., not used for stimulus triggering). The spike discrimination parameters are determined empirically by the user prior to the experiment and programmed into the chip. As stated previously, the flexibility in adjusting the spike-stimulus time delay is a critical system feature, because the relative timing of spikes and stimulus is known to influence conditioning effects in activity-dependent microstimulation in an intact nervous system [36].

C. Microstimulating Back-End

Fig. 4 shows the circuit schematic of the microstimulating back-end in each module, incorporating four identical stimulation channels with pMOS and nMOS current sources to generate

the anodic and cathodic current pulses, respectively, in biphasic stimulation. The stimulation channels also include a dedicated digital control unit for stimulus timing management as well as a 1.5-to-5 V signal level shifter, and share a 6-bit current-based digital-to-analog converter (DAC) and biasing circuitry available in each module.

To maintain a constant stimulus current regardless of the site and tissue impedances, current sources with high output impedance are required [37]. In this work, two feedback amplifiers, $A_{1,2}$, are used to boost the output impedance of the two current sources by maintaining a fixed voltage across drain-source of M_1 and M_4 [38]. Two compensation capacitors at the gates of M_2 (350 fF) and M_5 (150 fF) limit the amplifiers' bandwidth for stability [39].

As stated in Section II, an on-chip dc-dc converter generates a 5-V supply from 1.5 V for the microstimulating back-end. Since the output voltage of the converter is not fully regulated, it is important that the stimulator output current be relatively insensitive to supply voltage variation. Cascode current source architecture with active feedback circuitry as employed herein inherently has low sensitivity to supply voltage variation due to its high output impedance. However, supply voltage variation can also induce output current change through the biasing

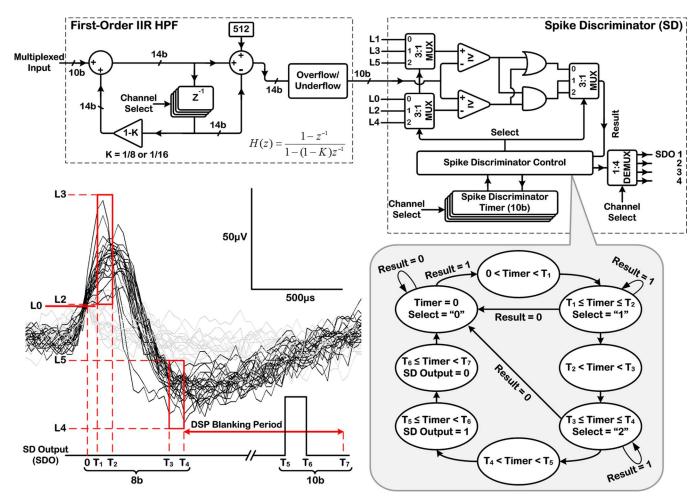


Fig. 3. Architecture of the DSP unit and operation of the time-amplitude window discriminator with neural spikes recorded wirelessly by our system from the cerebral cortex of a rat's brain. The flow chart for spike discrimination algorithm is also shown. Negative threshold level L_1 (not shown in bottom left) is used in the algorithm to discriminate waveforms with reverse polarity (i.e., negative-going initial portion), if necessary.

circuitry. To address this issue for the pMOS current source, M_{b5} is stacked on top of the nMOS current mirror in the current source biasing circuitry to increase the impedance seen from V_{bp} to ground. It should be noted that V_{bn} and V_{dn} for the nMOS current source are generated with circuitry powered by 1.5 V and therefore less susceptible to supply variation.

As previously described in [38], when source-drain voltage of M_5 in the pMOS current source is high (i.e., $|V_{ds}| > \sim 3 \text{ V}$), a significant portion of the transistor drain current can flow into its bulk due to impact ionization effect [40], greatly degrading the output impedance of the current source. This can be solved by connecting the bulk and source of M_5 together, as also seen in Fig. 4, allowing its drain-bulk current to also flow through M_4 and be controlled by the active feedback circuitry. However, this solution cannot be applied in a standard n-well CMOS process to address the same concern for M_2 in the nMOS current source.

Loizou and colleagues have proposed a 3-transistor MOS structure stacked on top of the current source with the purpose of limiting the drain-source voltage across each transistor to a reasonably small value for avoiding hot-carrier effects when the stimulator output voltage is near the supply rail [41]. In this work, this issue is addressed by adding $M_{\rm s1-5}$ to the nMOS current source in each stimulation channel, applying a voltage of $\sim (V_{\rm DD}/2 + V_{GS3})$ to the gate of $M_{\rm 3}$, instead of $V_{\rm DD}$, during the cathodic phase. In the worst-case scenario of the stimulator

output voltage being near the supply rail, the drain-source voltage of M_2 and M_3 is $\sim V_{\rm DD}/2$, not high enough to cause significant drain-bulk current in either transistor. When the output voltage decreases below $V_{\rm DD}/2$, M_3 enters the triode region with a small drain-source voltage, which does not adversely affect voltage compliance.

The microstimulator can be programmed to generate two different types of stimulus current pulses. With timing parameters $T_{1,2,3}$ set to be equal to T_{Anodic} (nominally 192 μ s), it generates a monophasic current pulse as shown in Fig. 4 [38]. To control the pulse amplitude, the 6-bit DAC generates a fraction (1/15) of the target output current, which is then amplified by the pMOS current source up to 94.5 μ A. This constant-current phase is followed by passive discharge to drain the accumulated charge on the stimulation site using a 2-bit programmable resistor (4.6 to 32 k Ω). During passive discharge, the current DAC and all biasing circuitry are powered down to reduce the static power consumption. In the monophasic mode, the node V_{body} and animal body are tied to the system ground, allowing output voltage headroom of 5 V.

The microstimulator can generate an asymmetric biphasic current pulse as well, as also shown in Fig. 4. In the leading anodic phase, a constant current up to 94.5 μ A is generated by the pMOS current source for duration T_1 followed by an interphase delay that lasts until T_2 . In the cathodic phase, a con-

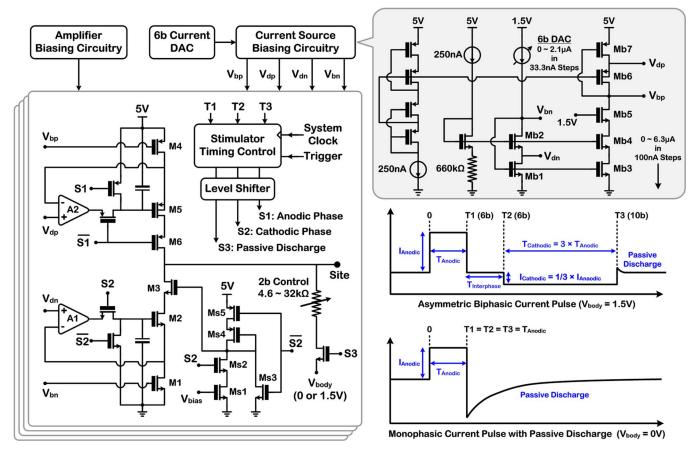


Fig. 4. Circuit schematic of the microstimulating back-end per module, comprising four identical stimulation channels with shared 6-bit current DAC and biasing circuitry. Each stimulation channel incorporates a timing control unit, a 1.5-to-5 V signal level shifter, and a pair of pMOS and nMOS current sources with boosted output impedance via active feedback. All transistors have a thick oxide, except for those in the timing control unit. The two stimulus current waveforms that can be generated by the stimulating back-end are also shown with corresponding time-amplitude parameters.

stant current up to 31.5 μ A (always 1/3 of the anodic phase amplitude) is generated by the nMOS current source that lasts until T_3 . With a 1-MHz system clock, T_1 and T_2 can be programmed up to 1.008 ms with a resolution of 16 μ s, whereas T_3 can be programmed up to 1.023 ms with a resolution of 1 μ s. For the asymmetric biphasic current pulse in Fig. 4 to be theoretically charge-balanced, T_3 should nominally be set equal to $3 \times T_1 + T_2$. In practice, however, the ratio of current levels in the anodic and cathodic phases can deviate from 1/3 due to process parameter variation and transistor mismatches. In that case, adjusting T_3 slightly around its nominal value can partially alleviate concerns associated with charge-imbalanced operation.

Passive discharge is also performed after each stimulation cycle to drain any residual charge left from charge mismatch between the anodic and cathodic phases. Further, in this proof-of-concept system with limited number of stimulation channels, external dc-blocking capacitors can also be placed in series with the electrode-tissue interface to prevent any net dc current flow into the tissue arising from charge imbalance or semiconductor failure. An alternative approach to ensure charge-balanced operation would be to employ *active* charge balancing based on monitoring the electrode voltage after each stimulation cycle and maintaining it within a safe predefined voltage range [42]. This technique would be more effective than passive discharge for high-frequency stimulation where there might not be adequate time for the discharge period to achieve sufficient charge

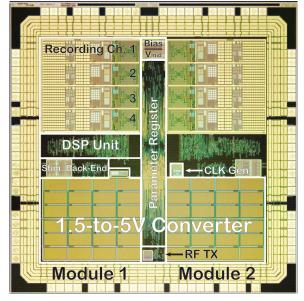


Fig. 5. Die micrograph of the 3.3 \times 3.3 mm 2 IC fabricated in 0.35- μ m 2P/4M CMOS.

balancing. However, this is not expected in *activity-dependent* stimulation given cortical neuronal firing rates of <150 spikes per second [43].

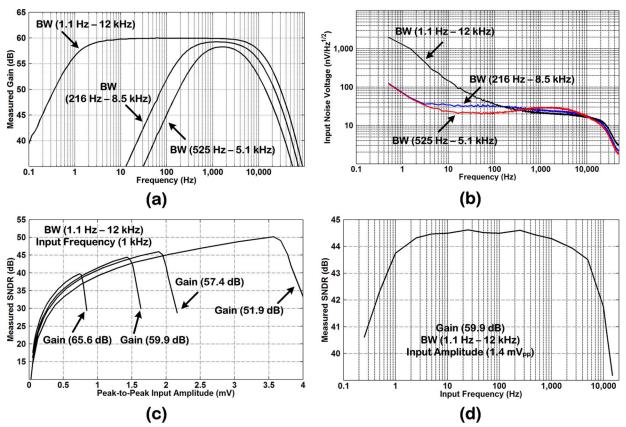


Fig. 6. Measured (a) frequency response and (b) input noise voltage of the analog recording front-end with different bandwidth settings. Measured SNDR of the entire recording front-end at the SAR ADC output versus (c) amplitude and (d) frequency of the input signal.

In the asymmetric biphasic mode, the node V_{body} and animal body are tied to 1.5 V, allowing output voltage headrooms of 3.5 V and 1.5 V for the anodic and cathodic phases, respectively. Therefore, the microstimulator has higher stimulus current drive capability in the monophasic mode due to higher output voltage headroom, but provides higher safety level in the biphasic mode by using a charge-balanced current waveform.

IV. MEASUREMENT RESULTS

A prototype chip was fabricated in 0.35- μ m two-poly four-metal (2P/4M) CMOS as shown in Fig. 5, measuring 3.3×3.3 mm² including the bonding pads. This section discusses the measurement results from benchtop characterization and acute biological experiments with rats.

A. Benchtop Characterization

The top plots in Fig. 6 depict the measured frequency response and input-referred noise voltage spectrum of the analog recording front-end for three different bandwidth settings and with the midband ac gain nominally set to 60 dB. The low cutoff frequency could be programmed from 1.1 to 525 Hz, whereas the high cutoff frequency could be adjusted in the range of 5.1 to 12 kHz. The midband ac gain was measured to be 51.9, 57.4, 59.9 and 65.6 dB when the $G_{\rm m}$ -C HPF was bypassed. The gain dropped by \sim 0.3 dB when the HPF was included in the signal path. With the bandwidth set to 1.1 Hz to 12 kHz, the thermal noise level was measured to be 21 nV/rtHz at 1 kHz. This level increased as the bandwidth decreased, because the LNA input transistors carried less current, increasing their thermal noise

contribution. With the same bandwidth setting, the total input noise voltage was measured to be 3.12 $\mu\rm V_{rms}$ by integrating the noise spectrum from 0.5 Hz to 50 kHz, indicating an increase of only 8.2% due to flicker noise. This led to a noise efficiency factor (NEF) of 2.68 for the LNA. The flicker noise corner frequency was measured to be < 90 Hz for all bandwidth settings, indicating that the $\rm G_m$ -C HPF could largely reduce the flicker noise contribution.

A sinusoidal signal with varying amplitude and frequency was then applied to the LNA input and the SNDR was measured at the ADC output. Fig. 6(c) shows the measured SNDR at 1 kHz versus input amplitude for the four available gain settings. Lower gain values provided higher input dynamic range, whereas higher gain values provided better signal resolution. Fig. 6(d) depicts the measured SNDR versus input frequency. In the frequency range of 100 Hz to 10 kHz for extracellular neural spikes, the recording front-end provided > 41.8 dB of accuracy.

The top plots in Fig. 7 depict the measured microstimulator output current versus its output voltage in the anodic and cathodic phases for four different DAC input codes. The stimulator output voltage could reach at least 4.68 V (going toward 5 V) and 150 mV (going toward 0 V) with a 5-V supply. To further examine the microstimulator functionality, it was interfaced with a silicon-substrate micromachined electrode with stimulus sites of iridium oxide (IrO). Fig. 7(c) and (d) depict the measured monophasic and asymmetric biphasic stimulus current waveforms delivered by the chip to saline via the microelectrode. The power consumption per channel was measured to be 3.5 μ W from 5 V, when the microstimulator generated

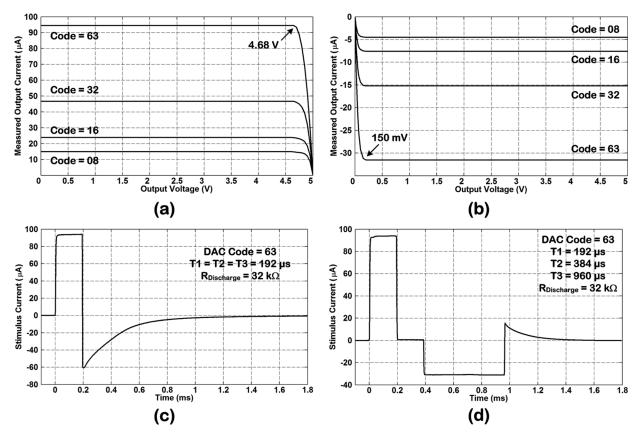


Fig. 7. Top: Measured microstimulator output current versus output voltage for four different DAC input codes in (a) anodic and (b) cathodic phases. Bottom: Measured (c) monophasic and (d) asymmetric biphasic stimulus current waveforms delivered by the chip to saline via a microelectrode. Passive discharge is also performed in each case using an on-chip 32-k Ω resistor.

the biphasic waveform in Fig. 7(d) at a rate of 33 Hz. Table I summarizes the measured performance of major circuitry, and Table II compares the system functionality and its measured performance with that in recent published work.

B. In Vivo Characterization

In the first biological experiment, a silicon microelectrode with recording sites of iridium was implanted in the somatosensory cortex of a rat's brain and externally interfaced with a single channel of the recording front-end on the chip. The left plot in Fig. 8 shows a 4-s snapshot of the extracellular neural spikes recorded wirelessly at the output of the ADC. In another test, a tungsten electrode with impedance value in the range of 50 to 100 k Ω was placed in the cortical motor area and externally interfaced with a single channel of the microstimulating back-end on the chip to stimulate the brain with a train of 13 monophasic current pulses (90 μ A, 192 μ s, 300 Hz). The right plot in Fig. 8 depicts the evoked electromyogram (EMG) signal recorded from the rat's neck muscle. The largest EMG spike appeared in <50 ms from the stimulation onset. The chip was then interfaced with the two implanted electrodes to demonstrate activity-dependent ICMS and programmed with the time-amplitude window discriminator parameters determined empirically prior to the experiment. In the first test, the recording and stimulation electrodes were each connected to a single channel of the front- and back-end on the chip, respectively. As the chip operated fully autonomously from a 1.5-V battery, it successfully delivered electrical stimuli to the cortical motor area triggered by neural spikes discriminated on the adjacent electrode in the somatosensory cortex, producing an artificial connection between the two brain regions. Fig. 9 shows the superimposed spike waveforms and corresponding stimulus artifacts from single-pulse stimulation (90 μ A, 192 μ s) with spike-stimulus delays of 5 and 20 ms. The stimulus artifact rejection mechanism in the recording front-end was de-activated in this test. Hence, artifacts of < 4 ms in duration were observed on the recording electrode.

Finally, while the stimulation electrode remained connected to a single channel of the back-end, the multisite recording electrode was connected to all four channels of the front-end in one module. The decision-making circuitry was programmed to trigger ICMS whenever neural activity would be present on any two or more data channels. Fig. 10 shows a 300-ms snapshot of the recorded data of each channel at the output of the digital HPF along with the corresponding SDO and the resulting stimulus trigger signal. Two large stimulus artifacts were distinguishable on each channel. The first stimulation was triggered by neural activity on channels 2 and 3, whereas the second was due to neural activity on channels 2 and 4. Some recorded spikes (marked by black arrows) were not discriminated by the DSP unit due to blanking of its operation after discriminating the spikes that immediately preceded them. This can be alleviated by reducing the time duration of DSP blanking.

TABLE I SUMMARY OF MEASURED PERFORMANCE

Record	ling Front-End	Microstimulating Back-End			
AC Gain @ 1 kHz	AC Gain @ 1 kHz 51.9, 57.4, 59.9, 65.6 dB		Anodic	Cathodic	
Low Cutoff Freq.	1.1 ~ 525 Hz	Stimulus	Asymmetric Biphasic &		
High Cutoff Freq.	5.1 ~ 12 kHz	Waveform	Monophasic w/ Passive Discha		
Total RMS Input	3.12 μV (BW of 12 kHz)	Output Current	0 ~ 94.5 μΑ	0 ~ 31.5 μΑ	
Noise Voltage	$3.42 \mu\text{V}$ (BW of 5.1kHz)	Current Pulsewidth	·		
NEF	2.68 (BW of 12 kHz)	Monophasic	0 ~ 1.008 ms	N/A	
NET	2.9 (BW of 5.1 kHz)	Biphasic	0 ~ 240 μs	0 ~ 720 μs	
CMRR @ 1 kHz	>56 dB	Output Impedance	~400 MΩ	~440 MΩ	
PSRR @ 1 kHz	>65 dB	DAC Resolution	6b		
Crosstalk	<-72 dB	DAC Linearity	<±1.6 LSB		
Max. Sampling Freq.	63 kS/s	Supply Sensitivity	-70.4 nA/V	-10.4 nA/V	
INL/DNL	$< \pm 0.8 \mathrm{LSB}$	Voltage Compliance			
ENOB	9.2b (f _{in} = 1 kHz, f _S = 35.7 kS/s) 9.1b (f _{in} = 1 kHz, f _S = 63 kS/s)	Monophasic Biphasic**	4.68 (of 5 V) 3.18 (of 3.5 V)	N/A 1.35 (of 1.5 V)	
Power Consumption*		Current Efficiency	95.6% (I _{out} = 94.5μ A)	N/A	
Amp-HPF	26.9 μW (BW of 12 kHz) 19.9 μW (BW of 5.1 kHz) 5.9 μW (f _{CLK} = 1 MHz)	1.5-to-5 V Converter			
SAR ADC		DC Output Voltage	4.86 ~ 5.35 V		
Digital Signal Processing Unit		Output Ripple	40 mV _{pp}		
HPF Cutoff Freq.	$366 / 756 \text{ Hz (f}_{CLK} = 1 \text{ MHz)}$	Max. Load Current	88 μ A ($V_{out} = 5.05 \text{ V}$)		
Power Consumption	$12.4 \mu W (f_{CLK} = 1 MHz)$	Power Efficiency	$31\% (V_{out} = 5.05 V)$		
Relaxation Oscillator		RF Transmitter			
		Comm. Scheme	FSK @ 433 MHz		
Output Frequency 420 kHz ~ 2.5 MHz		Received Power @	-55 dBm (18-cm monopole		
Supply Sensitivity	-60 kHz/V	1 m	TX and RX Antennae)		
Power Consumption	20.8 μ W (f _{CLK} = 1 MHz)	Power Consumption	200 μW		

Total Power for Two Modules w/o RF TX (Stimulation Rate = 33 Hz, Recording BW = 525 Hz \sim 5.1 kHz) = 375 μ W

TABLE II
COMPARISON OF SYSTEM FUNCTIONALITY AND MEASURED PERFORMANCE

	This Work	[13]	[14]	[17]	[19]
System Functionality	Neural Spike Recording; Neural Signal Processing; Neurostimulation; Power Conditioning	Neural Recording (Spikes & LFP)	Neural Recording (Spikes & EEG)	Neural Stimulation	Neural Stimulation
Supply Voltage	1.5 V (Battery Powered)	2.8 V	±2.5 V	5 V (Wirelessly Powered)	3.3 V (Logic) 10 V (Stimulation)
RMS Input Noise Voltage	3.12 μV (0.5 Hz – 50 kHz)	3.06 μV (10 Hz – 98 kHz)	2.2 µV (0.5 Hz – 50 kHz)	_	_
Power Consumption*	26.9 μW (BW of 12 kHz)	7.56 µW (BW of 5.32 kHz)	80 μW (BW of 7.2 kHz)	_	_
NEF	2.68	2.67	4	_	_
Stimulus Output Range/Resolution	0 – 94.5 μΑ/6b	_	_	1 – 255 μA/8b	0 – 735 μΑ
Stimulus Waveform	Asymmetric Biphasic & Monophasic w/ Passive Discharge	-	_	Symmetric Biphasic	Asymmetric Biphasic
Charge Balancing	Passive Discharge & 220-nF Blocking Capacitor	-	_	Charge Recovery w/ Active Circuit	100-nF Blocking Capacitor
Technology	0.35 μm CMOS	0.5 μm CMOS	1.5 μm CMOS	0.6 μm BiCMOS	0.35 μm CMOS
Experimental Paradigm	In Vivo	In Vivo	In Vivo	In Vivo	Benchtop
Publication Year	2011, JSSC	2007, TBCAS	2003, JSSC	2009, TBCAS	2008, TBCAS

^{*} Per recording channel

^{*} Per channel ** See Section III-C.

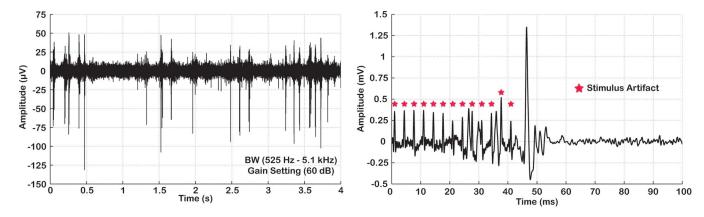


Fig. 8. In vivo demonstration of recording and stimulation functionalities of the chip in an anesthetized rat. Left – Extracellular neural spikes recorded wirelessly from the somatosensory cortex of the brain. The data are shown after linear-phase offline filtering (500 Hz to 4 kHz) with spike amplitudes referred to the input. Right – Measured EMG signal from the rat's neck muscle evoked by chip microstimulation of the cortical motor area with a train of 13 monophasic current pulses. The EMG signal is recorded by commercial electrophysiology equipment.

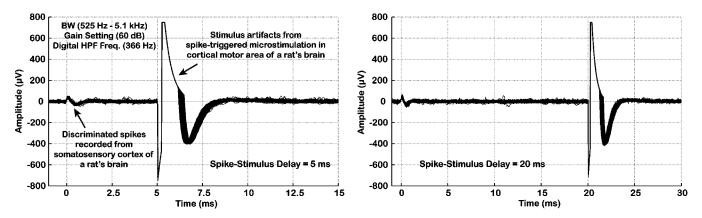


Fig. 9. Stimulation on one microelectrode in the cortical motor area of an anesthetized rat triggered by neural spikes discriminated on an adjacent microelectrode in the somatosensory cortex with spike-stimulus delays of 5 and 20 ms. Stimulus artifacts of < 4 ms in duration are observed on the recording electrode. The data are recorded wirelessly with the amplitude levels referred to the input.

As seen in Figs. 9 and 10, stimulus artifacts are used in this work to show that stimulation is occurring in vivo. However, to address the concern of artifacts false-triggering the microstimulator, a time-amplitude window discriminator is used (as opposed to simple spike thresholding) to discriminate between neural spikes and stimulus artifacts by proper adjustment of time-amplitude window parameters. Blanking the DSP operation after spike discrimination also prevents false-triggering the stimulator by the incoming stimulus artifact. Another concern related to stimulus artifacts is that any spike activity that occurs within the duration of the artifact cannot be detected by the system for triggering purposes. This is less of a concern for activity-dependent microstimulation, since we do not foresee a need to use every single spike in the system to entrain a neuronal population to another group of neurons. Nonetheless, we are developing an integrated signal-processing solution for real-time stimulus artifact rejection, which affords to retain signal information during each stimulation cycle [44], [45]. Such capability could be added to the DSP unit in the future.

V. DISCUSSION

A. LNA Noise Analysis

This section presents a noise analysis for the selected LNA topology and obtains a minimum NEF for a practical design

given our technology parameters and supply voltage. A similar discussion was presented in [13] for a slightly different LNA architecture. Fig. 11 shows a simplified schematic of the LNA and its core OTA for this analysis. We also model the transistor thermal noise current as

$$\overline{i_n^2} = 4KT \times \gamma \times q_m \tag{1}$$

where K is the Boltzmann's constant, T is the absolute temperature, g_m is the transistor transconductance, and γ is 2/3 for a transistor operating in strong inversion (above-threshold) and $1/(2\kappa)$ for a transistor operating in weak inversion (subthreshold) in which κ is the subthreshold gate coupling coefficient with a typical value of 0.7 [40]. The transistor transconductance can be estimated as

$$g_m \approx \frac{\kappa \times I_D}{V_{th}}$$
 Weak inversion (subthreshold)
 $g_m \approx \frac{2 \times I_D}{V_{OD}}$ Strong inversion (above-threshold) (2)

where V_{th} is the thermal voltage, I_D is the transistor drain current, and V_{OD} is the transistor overdrive voltage. Assuming that the overall transconductance of the OTA, G_m , is nearly equal

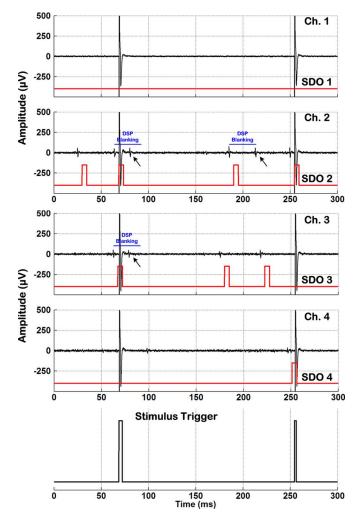


Fig. 10. Stimulation on one microelectrode in the cortical motor area of an anesthetized rat triggered by neural spikes discriminated on multiple recording sites of an adjacent microelectrode. The chip is programmed to trigger ICMS when neural activity is discriminated on any two or more channels of the recording front-end. The timing parameters T_5 , T_6 and T_7 in the DSP unit are set to 5, 10 and 28 ms, respectively. Arrows point to neural spikes on channels 2 and 3 that are not discriminated by the system due to blanking of the DSP operation.

to that of transistor M_1 , the input-referred noise voltage of the OTA can be estimated as

$$\overline{V_{n,OTA}^2} = \frac{1}{g_{m1}^2} \left(\frac{4KT}{\kappa} \times g_{m1} + \frac{16KT}{3} \times g_{m3} + \frac{16KT}{3} \times g_{m7} \right). \tag{3}$$

To minimize the OTA input noise voltage, $M_{1,2}$ operate in subthreshold to maximize g_{m1} for a given current level, whereas $M_{3,4,7,8}$ operate in strong inversion to reduce their transconductances. Moreover, the drain current of $M_{1,2}$ is selected to be much larger than that of $M_{7,8}$ in the folded branch. This current scaling scheme helps improve the OTA noise performance by further increasing g_{m1} and reducing $g_{m7,8}$. Therefore, neglecting the noise contribution by $M_{7,8}$, (3) can be simplified as

$$\overline{V_{n,OTA}^2} = \frac{1}{g_{m1}} \times \frac{4KT}{\kappa} \left(1 + \frac{8}{3} \times \frac{V_{th}}{V_{OD3}} \times (\beta + 1) \right) \quad (4)$$

where $\beta=I_7/I_1$ is the current scaling factor between the input differential pair and the folded branch. Equation (4) suggests that V_{OD3} should be increased to reduce the OTA input noise. However, increasing V_{OD3} means the drain-source voltage of $M_{3,4}$ should be increased too, limiting the OTA output voltage swing. For a peak-to-peak voltage swing (V_{Swing}) of 0.5 V at the OTA output, an upper limit of 0.5 V can be found for V_{OD3} given a supply voltage of 1.5 V. It should be noted that V_{Swing} of 0.5 V allows the LNA to handle input signals as large as $\sim \pm 6$ mV, in case LFPs or other low-frequency artifacts would also be present at the input. In this work, $M_{3,4}$ are sized for V_{OD3} of 0.35 V instead to ensure that they do not enter the triode region in the presence of process parameter variation.

According to (4), the OTA input noise voltage also depends on β , indicating that the current scaling factor should be selected judiciously. To that end, we next investigate the effect of β on the overall OTA transconductance, G_m . The G_m of a folded-cascode OTA with a similar current scaling scheme is thoroughly analyzed in [13]. In a similar way, the analysis of the proposed circuit reveals the G_m to be

$$G_m \approx \frac{g_{m5}}{g_{ds1} + g_{ds3} + g_{m5}} \times g_{m1}$$
 (5)

where g_{ds} is the transistor drain-source conductance. Since the channel length of $M_{3,4}$ is selected to be much longer than that of $M_{1,2}$, g_{ds3} is much smaller than g_{ds1} , which yields

$$\frac{G_m}{g_{m1}} \approx \frac{1}{1 + \frac{V_{th}}{\kappa \times V_{A1} \times \beta}} \tag{6}$$

where V_{A1} is the early voltage for M_1 (i.e., $1/\lambda_1$) and M_5 is sized to operate in subthreshold. This equation is also plotted in Fig. 11 with a value of 20 V for V_{A1} . According to (4), reducing β helps reduce the OTA input noise voltage, but β cannot be reduced arbitrarily to ensure that G_m is not degraded considerably (i.e., $\beta \geq 0.05$ from Fig. 11).

Another important consideration in selecting β is the sensitivity of bias currents to transistor mismatches in the OTA and associated biasing circuitry. Assume that ΔI_1 , ΔI_3 and ΔI_7 are bias current deviations from their corresponding nominal values due to transistor mismatches. If ΔI_1 and ΔI_3 are assumed to be uncorrelated, it can be shown that

$$\frac{\Delta I_7}{I_7} = \sqrt{\frac{\Delta I_1^2 + \Delta I_3^2}{I_7^2}}
= \frac{1}{\beta} \times \sqrt{\left(\frac{\Delta I_1}{I_1}\right)^2 + (\beta + 1)^2 \times \left(\frac{\Delta I_3}{I_3}\right)^2}.$$
(7)

For example, for a β value of 0.05, a 2% variation in I_1 and I_3 causes a 58% variation in I_7 , which might adversely affect the OTA operation and degrade its transconductance. In this work, the minimum value of β (when LNA bandwidth is maximum) is selected to be 0.091 for which G_m is 98% of g_{m1} , and 2% variation in I_1 and I_3 causes only 32.5% variation in I_7 . Table III tabulates the dimension, current level, and operating condition of each transistor pair in the OTA for optimum noise performance with the maximum bandwidth setting.

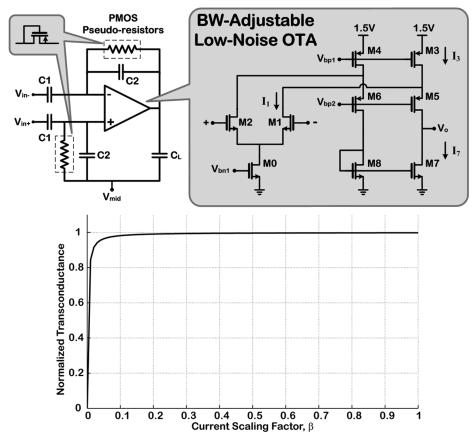


Fig. 11. Simplified schematic of the LNA and its core OTA for noise analysis (top) and a plot of the normalized OTA transconductance G_m/g_{m1} versus the current scaling factor β (bottom).

TABLE III
DIMENSION, CURRENT LEVEL, AND OPERATING CONDITION OF OTA TRANSISTORS

Transistor	Dimension (µm)	Current Level (µA)	Simulated g _m (μS)	Inversion Coefficient*	Operating Condition
$M_{1,2}$	800/2	2.75	76	0.021	subthreshold
$M_{3,4}$	20/20	3.00	16	26.8	above-threshold
$M_{5,6}$	80/1.3	0.25	6.9	0.036	subthreshold
$M_{7,8}$	4/40	0.25	2.5	7.6	above-threshold

^{*} $\mu_n C_{OX} = 170 \ \mu A/V^2$, $\mu_p C_{OX} = 58 \ \mu A/V^2$

To compute the NEF, we should note that the input-referred noise voltage of the LNA is almost equal to that of the OTA given that the parasitic gate capacitance at the input terminals of the OTA is typically much smaller than $C_{1,2}$. The NEF can be calculated according to [46]

NEF =
$$V_{ni,\text{rms}} \times \sqrt{\frac{2 \times I_{\text{total}}}{\pi \times V_{th} \times 4KT \times BW}}$$
 (8)

where $V_{ni,\rm rms}$ is the rms input noise voltage of the LNA, $I_{\rm total}$ is the total supply current, and BW is the 3-dB bandwidth of the amplifier. Finally, assuming that the LNA has a single dominant pole in its frequency response and noting that $I_{\rm total}=2\times I_3$, combining (4) and (8) yields

NEF =
$$\sqrt{\frac{2 \times (\beta + 1)}{\kappa^2} \times \left(1 + \frac{8}{3} \times \frac{V_{th}}{V_{OD3}} \times (\beta + 1)\right)}$$
 (9)

which results in an NEF of 2.33 for β of 0.091 and V_{OD3} of 0.35 V. Hence, the NEF of 2.68 derived from measured performance of the LNA (when set for maximum bandwidth) is in

good agreement with this analysis. This argument also shows that reducing the supply voltage can adversely impact the OTA noise performance for the same output voltage swing, making it challenging to reduce $V_{\rm DD}$ below 1.5 V in this architecture.

B. Future Considerations in System Design

Future applications of the proposed activity-dependent ICMS system might require 16, 32 or even higher number of recording and stimulating channels, resulting in higher power consumption and larger silicon area. Fig. 12 shows a breakdown of the area and power consumption for the entire SoC, including both modules. The core area of the SoC excluding I/O pads is $2.6 \times 2.6 \text{ mm}^2$, of which 34% is occupied by the 1.5-to-5-V converter. In this work, the converter can provide a maximum dc load current of $\sim 88~\mu\text{A}$ for an output voltage of 5.05 V, which allows an average stimulus rate of >500 Hz for simultaneous stimulation on all eight channels with maximum stimulus current, given that the microstimulator dissipates 3.5 μW per channel for such stimulation at a rate of 33 Hz.

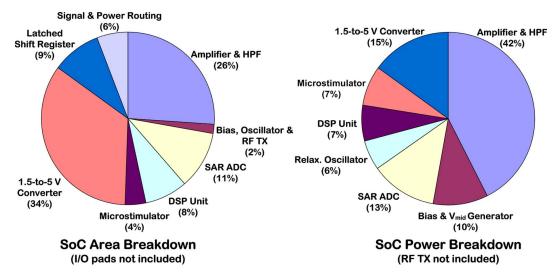


Fig. 12. SoC area and power breakdown.

However, with cortical neuronal firing rates of <150 spikes per second [43], the silicon area of the proposed converter can be reduced by $\sim63\%$.

The power pie-chart is generated assuming biphasic stimulation at a rate of 33 Hz (anodic: 94.5 μ A, 192 μ s; cathodic: $31.5 \,\mu\text{A}$, $576 \,\mu\text{s}$) and recording bandwidth of $525 \,\text{Hz}$ to $5.1 \,\text{kHz}$. Excluding the FSK transmitter, the total power consumption is measured to be 375 μ W for two modules with the analog recording front-end being the most power-hungry circuit block. Based on simulation results, if we reduce the LNA bias currents to half their current values (all transistors and capacitors should also be sized down accordingly to maintain the same bandwidth), we can save up to 29% and 20% in silicon area and power consumption of the analog recording front-end, respectively. Although this would increase the LNA total input noise by $\sim 41\%$ to 4.8 $\mu V_{\rm rms}$, it is still less than the background noise of the recording site (5 to 10 μ V) [47]. Another effective approach to reduce the front-end power consumption and silicon area is to reduce the SAR ADC resolution to 9 bits. In the proposed design, the input-referred quantization noise of the SAR ADC $(V_{\rm DD}/(\sqrt{12} \times {\rm Gain} \times 2^{\rm ENOB}))$ is $\sim 0.8 \ \mu {\rm V_{rms}}$ for a nominal gain of 60 dB, which is much less than that of the LNA (\sim 3 μ V_{rms}). Therefore, reducing the SAR ADC resolution by one bit would not degrade the performance considerably. Further, the power consumption of the secondary amplifier (\sim 17% of the total system power) driving the capacitive network of the ADC can be significantly reduced as well by decreasing the ADC resolution.

VI. CONCLUSION

Brain-machine-brain interfaces (BMBIs) are a new generation of neuroprostheses that use information extracted from brain neural activity for titration control of neuromodulation in real time, potentially enhancing their clinical efficacy in restoring a lost motor or sensory function in the disabled. In this paper, we reported on the design, implementation, performance characterization, and *in vivo* testing of a battery-powered IC for activity-dependent ICMS. The single-chip

system integrated recording and digitization front-end, neural signal processing unit, 1.5-to-5 V converter, and microstimulating back-end to convert extracellular neural spikes recorded from the somatosensory cortex of a rat's brain to electrical stimuli delivered to the cortical motor area in real time with user-adjustable spike-stimulus time delay, when operating autonomously from a single battery. This system is envisioned to offer a novel *device-based* approach to repairing damaged neural pathways directly within the brain with future rehabilitative applications in behavioral recovery from traumatic brain injury (TBI) [48].

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